

IDENTIFICATION OF NITROGEN-FIXING BACTERIA FROM THE RECLAIMED SMELTER WASTELAND USING 16S rDNA SEQUENCING METHOD AND GENOTYPING OF STRAINS USING PCR-MP.

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PIEKARY ŚLĄSKIE

Smelter wastelands with extreme contents of zinc, lead, cadmium and arsenic constitute a serious problem in the area of Upper Silesia. Soil contamination with metals can cause toxicity to plants and soil microorganisms, limiting the land ability to perform basic functions.

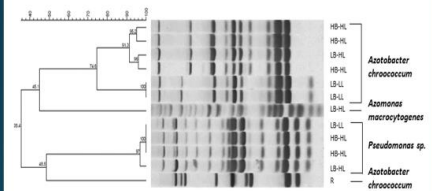
The activity of N-binding bacteria is extremely important for the development of the plant cover in these areas. In 2016, samples taken from a long-term field experiment (reclamation in 1997) at a waste heap in Piekary Śląskie were collected from a depth of 0-20 cm in three replications.

The material was collected from 6 transects representing different combinations of sewage sludge and by-product lime rates. The method of plate dilutions was used to determine the total number of Azotobacter which was expressed as the number of colonies (CFU) in 1 gram of air dried soil matter. Then bacterial species identification was performed by 16S rDNA sequencing and PCR genotyping of strains.

As a result of the genotyping of the diazotroph strains, 7 different electrophoretic profiles were obtained.

The dimension of the whole experimental site was 40m x 100m. Six different treatments of the top layer. Were applied in the plot experiment and compared with the untreated control:

- LB – lower biosolids (150 t ha⁻¹),
- HB – higher biosolids rate (300 t ha⁻¹),
- LB-LL – lower biosolids (150 t ha⁻¹) and lower lime (100 t ha⁻¹) rates,
- HB-LL – higher biosolids (300 t ha⁻¹) and lower lime (100 t ha⁻¹) rates,
- LB-HL – lower biosolids (150 t ha⁻¹) and higher lime 1000 t ha⁻¹ rates,
- HB-HL – higher biosolids (300 t ha⁻¹) and higher lime (1000 t ha⁻¹) rates.



	H ₂ O - extractable						Total content									
	Cd		Zn		Pb		Cd		Zn		As		Hg			
	mean	STD	mean	STD	mean	STD	mean	STD	mean	STD	mean	STD	mean	STD		
Control	0.016 ^{ns}	0.010	0.07 ^a	0.04	0.07 ^a	0.04	163 ^{ns}	114	4876 ^{ns}	2953	19859 ^a	9781	1286 ^a	195	0.52 ^a	0.45
LB	0.021 ^{ns}	0.015	0.74 ^{ab}	0.29	0.41 ^{ab}	0.13	607 ^a	412	1639 ^a	8164	24040 ^{ab}	7003	1219 ^a	162	2.15 ^{ab}	0.91
HB	0.027 ^a	0.013	1.49 ^b	0.49	0.76 ^b	0.36	242 ^a	109	6351 ^{ab}	2840	7613 ^a	2525	273 ^a	111	2.70 ^b	0.29
LB-LL	0.009 ^b	0.002	0.95 ^{ab}	0.25	0.28 ^b	0.06	334 ^a	92	9483 ^{ab}	2761	26274 ^{ab}	8056	672 ^a	5	2.09 ^{ab}	0.56
HB-LL	0.013 ^{ns}	0.005	1.19 ^{ab}	0.47	0.32 ^{bc}	0.10	224 ^a	14	6503 ^{ab}	472	17521 ^a	3957	677 ^a	200	1.71 ^b	0.47
LB-HL	0.002 ^a	0.000	0.18 ^b	0.05	0.64 ^{ab}	0.02	134 ^a	11	3052 ^a	396	16203 ^a	102	213 ^a	1	0.98 ^b	0.23
HB-HL	0.003 ^a	0.001	0.82 ^{ab}	0.48	0.68 ^{ab}	0.03	95 ^a	4	2303 ^a	175	13425 ^a	456	173 ^a	19	1.86 ^b	0.21

Microbiological and biochemical analysis
In order to characterize biochemical activity of the amended wasteland, activities of three enzymes (dehydrogenases, acidic and alkaline phosphatases) were measured using standard protocols (Tabatabai, 1994). The determination of dehydrogenases was performed according to Casida et al. (1964) by the colorimetric method, using TTC (triphenyltetrazole chloride) as a substrate, after 24 hours incubation at 37°C. Alkaline and acid phosphatase activities were measured by colorimetric method using PNP (sodium p-nitrophenylphosphate) after 1 hour incubation at 37 ° C at 410 nm wavelength (Tabatabai and Bremner 1969).

	Dehydrogenases µg TPF g soil DM ⁻¹ h ⁻¹		Alkaline phosphatase µg p-nitrophenyl g soil DM ⁻¹ h ⁻¹		Acidic phosphatase µg p-nitrophenyl g soil DM ⁻¹ h ⁻¹	
	mean	STD	mean	STD	mean	STD
	Control	0.325 ^{ns}	0.04	27.0 ^{ns}	18.4	26.0 ^{ns}
I	0.09 ^{ns}	0.02	22.1 ^a	6.0	39.4 ^a	10.3
II	0.16 ^{ns}	0.00	36.5 ^b	0.9	67.8 ^a	3.5
III	0.22 ^a	0.05	52.5 ^b	11.3	86.8 ^b	18.8
IV	0.27 ^{ab}	0.08	71.0 ^{ab}	29.6	118.9 ^b	38.8
V	0.20 ^{ab}	0.03	47.4 ^a	5.5	85.2 ^a	8.2
VI	0.40 ^b	0.05	91.3 ^b	8.0	154.7 ^b	18.0

The total count of bacteria and actinobacteria (Wallace and Lockhead, 1950), ammonification bacteria (Rodina, 1968), bacteria of *Azotobacter* (Fenglerova, 1965) and total count of fungi (Martin 1950) were determined by the plate dilution method. The plates were incubated in temperature 28°C and the number of colonies was counted after 3 – 5 days of growth. All biochemical and microbiological measurements were done in triplicate.

	Total number of bacteria		Total number of bacteria <i>Azotobacter</i>		Total number of ammonification bacteria		Total number of fungi	
	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	10 ⁶ CFU g ⁻¹ d.m. of soil	
Control	2.35 ^a	1.14	0.00 ^a	0.00	0.25 ^a	0.15	12.76 ^a	3.86
I	6.32 ^b	2.60	0.00 ^a	0.00	1.80 ^{ab}	1.16	12.31 ^a	2.99
II	42.93 ^c	17.02	0.00 ^a	0.00	0.81 ^a	0.59	121.44 ^b	21.40
III	100.38 ^d	63.57	207.64 ^{bc}	102.60	2.97 ^b	0.28	62.52 ^{ab}	45.48
IV	95.38 ^d	35.58	180.61 ^{bc}	76.91	5.16 ^{bc}	4.67	44.47 ^b	16.28
V	47.63 ^c	13.81	221.17 ^{bc}	19.20	5.94 ^{bc}	3.77	55.70 ^b	27.40
VI	155.43 ^e	34.40	111.91 ^b	15.14	9.44 ^c	2.56	73.61 ^b	37.20